

# Package ‘pmd’

October 14, 2022

**Type** Package

**Title** Paired Mass Distance Analysis for GC/LC-MS Based Non-Targeted Analysis and Reactomics Analysis

**Version** 0.2.1

**Date** 2021-01-20

**Maintainer** Miao YU <yufreecas@gmail.com>

**Description** Paired mass distance (PMD) analysis proposed in Yu, Olkowicz and Pawliszyn (2018) <[doi:10.1016/j.aca.2018.10.062](https://doi.org/10.1016/j.aca.2018.10.062)> for gas/liquid chromatography–mass spectrometry (GC/LC-MS) based non-targeted analysis. PMD analysis including GlobalStd algorithm and structure/reaction directed analysis. GlobalStd algorithm could found independent peaks in m/z-retention time profiles based on retention time hierarchical cluster analysis and frequency analysis of paired mass distances within retention time groups. Structure directed analysis could be used to find potential relationship among those independent peaks in different retention time groups based on frequency of paired mass distances. Reactomics analysis could also be performed to build PMD network, assign sources and make biomarker reaction discovery. GUIs for PMD analysis is also included as 'shiny' applications.

**URL** <https://yufree.github.io/pmd/>

**BugReports** <https://github.com/yufree/pmd/issues>

**License** GPL-2

**Encoding** UTF-8

**LazyData** true

**Suggests** knitr, shiny, rmarkdown, MSnbase

**VignetteBuilder** knitr

**biocViews**

**Depends** R (>= 3.5.0)

**Imports** RColorBrewer, stats, graphics, utils, data.table, igraph, enviGCMS

**RoxygenNote** 7.1.1

**NeedsCompilation** no

**Author** Miao YU [aut, cre] (<<https://orcid.org/0000-0002-2804-6014>>)

Repository CRAN

Date/Publication 2021-01-21 23:20:08 UTC

## R topics documented:

getcda . . . . .	2
getchain . . . . .	3
getcluster . . . . .	4
getcorcluster . . . . .	5
getms2pmd . . . . .	5
getmspmd . . . . .	6
getpaired . . . . .	6
getpmd . . . . .	7
getposneg . . . . .	8
getrda . . . . .	9
getreact . . . . .	10
getsda . . . . .	11
getstd . . . . .	12
gettarget . . . . .	13
globalstd . . . . .	13
hmdb . . . . .	14
kegrall . . . . .	15
omics . . . . .	15
pcasf . . . . .	16
ploten . . . . .	17
plotpaired . . . . .	17
plotrtg . . . . .	18
plotsda . . . . .	18
plotstd . . . . .	19
plotstdrt . . . . .	19
plotstdsda . . . . .	20
pmdanno . . . . .	21
runPMD . . . . .	21
runPMDnet . . . . .	21
sda . . . . .	22
spmein vivo . . . . .	22
<b>Index</b>	<b>23</b>

---

getcda	<i>Perform correlation directed analysis for peaks list.</i>
--------	--

---

## Description

Perform correlation directed analysis for peaks list.

**Usage**

```
getcda(list, corcutoff = 0.9, rtcutoff = 10, accuracy = 4)
```

**Arguments**

list	a list with mzrt profile
corcutoff	cutoff of the correlation coefficient, default NULL
rtcutoff	cutoff of the distances in retention time hierarchical clustering analysis, default 10
accuracy	measured mass or mass to charge ratio in digits, default 4

**Value**

list with correlation directed analysis results

**See Also**

[getsda](#), [getrda](#)

**Examples**

```
data(spmein vivo)
cluster <- getcorcluster(spmein vivo)
cbp <- enviGMS::getfilter(cluster, rowindex = cluster$stdmassindex2)
cda <- getcda(cbp)
```

---

getchain

*Get reaction chain for specific mass to charge ratio*

---

**Description**

Get reaction chain for specific mass to charge ratio

**Usage**

```
getchain(
  list,
  diff,
  mass,
  digits = 2,
  accuracy = 4,
  rtcutoff = 10,
  corcutoff = 0.6,
  ppm = 25
)
```

**Arguments**

list	a list with mzrt profile
diff	paired mass distance(s) of interests
mass	a specific mass for known compound or a vector of masses. You could also input formula for certain compounds
digits	mass or mass to charge ratio accuracy for pmd, default 2
accuracy	measured mass or mass to charge ratio in digits, default 4
rtcutoff	cutoff of the distances in retention time hierarchical clustering analysis, default 10
corcutoff	cutoff of the correlation coefficient, default NULL
ppm	all the peaks within this mass accuracy as seed mass or formula

**Value**

a list with mzrt profile and reaction chain dataframe

**Examples**

```
data(spmein vivo)
# check metabolites of C18H39NO
pmd <- getchain(spmein vivo,diff = c(2.02,14.02,15.99),mass = 286.3101)
```

---

getcluster

*Get Pseudo-Spectrum as peaks cluster based on pmd analysis.*

---

**Description**

Get Pseudo-Spectrum as peaks cluster based on pmd analysis.

**Usage**

```
getcluster(list, corcutoff = NULL, accuracy = 4)
```

**Arguments**

list	a list from getstd function
corcutoff	cutoff of the correlation coefficient, default NULL
accuracy	measured mass or mass to charge ratio in digits, default 4

**Value**

list with Pseudo-Spectrum index

**See Also**

[getpaired](#),[getstd](#),[plotstd](#)

**Examples**

```
data(spmein vivo)
re <- getpaired(spmein vivo)
re <- getstd(re)
cluster <- getcluster(re)
```

---

getcorcluster	<i>Get Pseudo-Spectrum as peaks cluster based on correlation analysis.</i>
---------------	--

---

**Description**

Get Pseudo-Spectrum as peaks cluster based on correlation analysis.

**Usage**

```
getcorcluster(list, corcutoff = 0.9, rtcutoff = 10, accuracy = 4)
```

**Arguments**

list	a list with peaks intensity
corcutoff	cutoff of the correlation coefficient, default 0.9
rtcutoff	cutoff of the distances in cluster, default 10
accuracy	measured mass or mass to charge ratio in digits, default 4

**Value**

list with Pseudo-Spectrum index

**Examples**

```
data(spmein vivo)
cluster <- getcorcluster(spmein vivo)
```

---

getms2pmd	<i>read in MSP file as list for ms/ms annotation</i>
-----------	--

---

**Description**

read in MSP file as list for ms/ms annotation

**Usage**

```
getms2pmd(file, digits = 2, icf = 10)
```

**Arguments**

file            the path to your MSP file  
 digits        mass or mass to charge ratio accuracy for pmd, default 2  
 icf            intensity cutoff, default 10 percentage

**Value**

list a list with MSP information for MS/MS annotation

---

getmspmd            *read in MSP file as list for EI-MS annotation*

---

**Description**

read in MSP file as list for EI-MS annotation

**Usage**

```
getmspmd(file, digits = 2, icf = 10)
```

**Arguments**

file            the path to your MSP file  
 digits        mass or mass to charge ratio accuracy for pmd, default 0  
 icf            intensity cutoff, default 10 percentage

**Value**

list a list with MSP information for EI-MS annotation

---

getpaired            *Filter ions/peaks based on retention time hierarchical clustering, paired mass distances(PMD) and PMD frequency analysis.*

---

**Description**

Filter ions/peaks based on retention time hierarchical clustering, paired mass distances(PMD) and PMD frequency analysis.

**Usage**

```
getpaired(list, rtcutoff = 10, ng = NULL, digits = 2, accuracy = 4)
```

**Arguments**

list	a list with mzrt profile
rtcutoff	cutoff of the distances in retention time hierarchical clustering analysis, default 10
ng	cutoff of global PMD's retention time group numbers, If ng = NULL, 20 percent of RT cluster will be used as ng, default NULL.
digits	mass or mass to charge ratio accuracy for pmd, default 2
accuracy	measured mass or mass to charge ratio in digits, default 4

**Value**

list with tentative isotope, multi-chargers, adducts, and neutral loss peaks' index, retention time clusters.

**See Also**

[getstd](#), [getsda](#), [plotpaired](#)

**Examples**

```
data(spmein vivo)
pmd <- getpaired(spmein vivo)
```

---

getpmd

*Get pmd for specific reaction*

---

**Description**

Get pmd for specific reaction

**Usage**

```
getpmd(list, pmd, rtcutoff = 10, corcutoff = NULL, digits = 2, accuracy = 4)
```

**Arguments**

list	a list with mzrt profile
pmd	a specific paired mass distance
rtcutoff	cutoff of the distances in retention time hierarchical clustering analysis, default 10
corcutoff	cutoff of the correlation coefficient, default NULL
digits	mass or mass to charge ratio accuracy for pmd, default 2
accuracy	measured mass or mass to charge ratio in digits, default 4

**Value**

list with paired peaks for specific pmd.

**See Also**

[getpaired](#), [getstd](#), [getsda](#), [getrda](#)

**Examples**

```
data(spmein vivo)
pmd <- getpmd(spmein vivo, pmd=15.99)
```

---

getposneg

*Link pos mode peak list with neg mode peak list by pmd.*

---

**Description**

Link pos mode peak list with neg mode peak list by pmd.

**Usage**

```
getposneg(pos, neg, pmd = 2.02, digits = 2)
```

**Arguments**

pos	a list with mzrt profile collected from positive mode.
neg	a list with mzrt profile collected from negative mode.
pmd	numeric or numeric vector
digits	mass or mass to charge ratio accuracy for pmd, default 2

**Value**

dataframe with filtered positive and negative peak list



---

getrda	<i>Perform structure/reaction directed analysis for mass only.</i>
--------	--

---

## Description

Perform structure/reaction directed analysis for mass only.

## Usage

```
getrda(mz, freqcutoff = 10, digits = 3, top = 20, formula = NULL)
```

## Arguments

mz	numeric vector for independent mass or mass to charge ratio. Mass to charge ratio from GlobalStd algorithm is suggested. Isomers would be excluded automatically
freqcutoff	pmd frequency cutoff for structures or reactions, default 10
digits	mass or mass to charge ratio accuracy for pmd, default 3
top	top n pmd frequency cutoff when the freqcutoff is too small for large data set
formula	vector for formula when you don't have mass or mass to charge ratio data

## Value

logical matrix with row as the same order of mz or formula and column as high frequency pmd group

## See Also

[getsda](#)

## Examples

```
data(spmein vivo)
pmd <- getpaired(spmein vivo)
std <- getstd(pmd)
sda <- getrda(spmein vivo$mz[std$stdmassindex])
```

---

 getreact

*Get quantitative paired peaks list for specific reaction/pmd*


---

### Description

Get quantitative paired peaks list for specific reaction/pmd

### Usage

```
getreact(
  list,
  pmd,
  rtcutoff = 10,
  digits = 2,
  accuracy = 4,
  ratio cv = 30,
  outlier = F,
  ...
)
```

### Arguments

list	a list with mzrt profile and data
pmd	a specific paired mass distances
rtcutoff	cutoff of the distances in retention time hierarchical clustering analysis, default 10
digits	mass or mass to charge ratio accuracy for pmd, default 2
accuracy	measured mass or mass to charge ratio in digits, default 4
ratio cv	ratio cv cutoff for quantitative paired peaks, default 30
outlier	logical, if true, outlier of ratio will be removed, default False.
...	other parameters for getpmd

### Value

list with quantitative paired peaks.

### See Also

[getpaired](#), [getstd](#), [getsda](#), [getrda](#), [getpmd](#),

### Examples

```
data(spmein vivo)
pmd <- getreact(spmein vivo, pmd=15.99)
```

---

getsda	<i>Perform structure/reaction directed analysis for peaks list.</i>
--------	---

---

**Description**

Perform structure/reaction directed analysis for peaks list.

**Usage**

```
getsda(  
  list,  
  rtcutoff = 10,  
  corcutoff = NULL,  
  digits = 2,  
  accuracy = 4,  
  freqcutoff = NULL  
)
```

**Arguments**

<code>list</code>	a list with mzrt profile
<code>rtcutoff</code>	cutoff of the distances in retention time hierarchical clustering analysis, default 10
<code>corcutoff</code>	cutoff of the correlation coefficient, default NULL
<code>digits</code>	mass or mass to charge ratio accuracy for pmd, default 2
<code>accuracy</code>	measured mass or mass to charge ratio in digits, default 4
<code>freqcutoff</code>	pmd frequency cutoff for structures or reactions, default NULL. This cutoff will be found by PMD network analysis when it is NULL.

**Value**

list with tentative isotope, adducts, and neutral loss peaks' index, retention time clusters.

**See Also**

[getpaired](#), [getstd](#), [plotpaired](#)

**Examples**

```
data(spmein vivo)  
pmd <- getpaired(spmein vivo)  
std <- getstd(pmd)  
sda <- getsda(std)
```

---

getstd	<i>Find the independent ions for each retention time hierarchical clustering based on PMD relationship within each retention time cluster and isotope and return the index of the std data for each retention time cluster.</i>
--------	---

---

### Description

Find the independent ions for each retention time hierarchical clustering based on PMD relationship within each retention time cluster and isotope and return the index of the std data for each retention time cluster.

### Usage

```
getstd(list, corcutoff = NULL, digits = 2, accuracy = 4)
```

### Arguments

list	a list from getpaired function
corcutoff	cutoff of the correlation coefficient, default NULL
digits	mass or mass to charge ratio accuracy for pmd, default 2
accuracy	measured mass or mass to charge ratio in digits, default 4

### Value

list with std mass index

### See Also

[getpaired](#), [getsda](#), [plotstd](#)

### Examples

```
data(spmein vivo)
pmd <- getpaired(spmein vivo)
std <- getstd(pmd)
```

---

gettarget	<i>Get multiple injections index for selected retention time</i>
-----------	--

---

**Description**

Get multiple injections index for selected retention time

**Usage**

```
gettarget(rt, drt = 10, n = 6)
```

**Arguments**

rt	retention time vector for peaks in seconds
drt	retention time drift for targeted analysis in seconds, default 10.
n	max ions numbers within retention time drift windows

**Value**

index for each injection

**Examples**

```
data(spmeinivo)
pmd <- getpaired(spmeinivo)
std <- getstd(pmd)
index <- gettarget(std$rt[std$stdmassindex])
table(index)
```

---

globalstd	<i>GlobalStd algorithm with structure/reaction directed analysis</i>
-----------	--

---

**Description**

GlobalStd algorithm with structure/reaction directed analysis

**Usage**

```
globalstd(
  list,
  rtcutoff = 10,
  ng = NULL,
  corcutoff = NULL,
  digits = 2,
  accuracy = 4,
  freqcutoff = NULL,
  sda = FALSE
)
```

**Arguments**

<code>list</code>	a peaks list with mass to charge, retention time and intensity data
<code>rtcutoff</code>	cutoff of the distances in cluster, default 10
<code>ng</code>	cutoff of global PMD's retention time group numbers, If <code>ng = NULL</code> , 20 percent of RT cluster will be used as <code>ng</code> , default <code>NULL</code> .
<code>corcutoff</code>	cutoff of the correlation coefficient, default <code>NULL</code>
<code>digits</code>	mass or mass to charge ratio accuracy for <code>pmd</code> , default 2
<code>accuracy</code>	measured mass or mass to charge ratio in digits, default 4
<code>freqcutoff</code>	<code>pmd</code> frequency cutoff for structures or reactions, default <code>NULL</code> . This cutoff will be found by PMD network analysis when it is <code>NULL</code> .
<code>sda</code>	logical, option to perform structure/reaction directed analysis, default <code>FALSE</code> .

**Value**

list with GlobalStd algorithm processed data.

**See Also**

[getpaired](#), [getstd](#), [getsda](#), [plotstd](#), [plotstdsda](#), [plotstdrt](#)

**Examples**

```
data(spmein vivo)
re <- globalstd(spmein vivo)
```

---

<code>hmdb</code>	<i>A dataframe containing HMDB with unique accurate mass <code>pmd</code> with three digits frequency larger than 1 and accuracy percentage larger than 0.9.</i>
-------------------	--

---

**Description**

A dataframe containing HMDB with unique accurate mass `pmd` with three digits frequency larger than 1 and accuracy percentage larger than 0.9.

**Usage**

```
data(hmdb)
```

**Format**

A dataframe with atoms numbers of C, H, O, N, P, S

**percentage** accuracy of atom numbers prediction

**pmd2** `pmd` with two digits

**pmd** `pmd` with three digits

---

keggrall	<i>A dataframe containing reaction related accurate mass pmd and related reaction formula with KEGG ID</i>
----------	--

---

**Description**

A dataframe containing reaction related accurate mass pmd and related reaction formula with KEGG ID

**Usage**

```
data(keggrall)
```

**Format**

A dataframe with KEGG reaction, their related pmd and atoms numbers of C, H, O, N, P, S

**ID** KEGG reaction ID

**pmd** pmd with three digits

---

omics	<i>A dataframe containing multiple reaction database ID and their related accurate mass pmd and related reactions</i>
-------	---

---

**Description**

A dataframe containing multiple reaction database ID and their related accurate mass pmd and related reactions

**Usage**

```
data(omics)
```

**Format**

A dataframe with reaction and their related pmd

**KEGG** KEGG reaction ID

**RHEA\_ID** RHEA\_ID

**DIRECTION** reaction direction

**MASTER\_ID** master reaction RHEA ID

**ec** ec reaction ID

**ecocyc** ecocyc reaction ID

**macie** macie reaction ID

**metacyc** metacyc reaction ID  
**reactome** reactome reaction ID  
**compounds** reaction related compounds  
**pmd** pmd with two digits  
**pmd2** pmd with three digits

---

pcasf

*Compare matrices using PCA similarity factor*

---

### Description

Compare matrices using PCA similarity factor

### Usage

```
pcasf(x, y, dim = NULL)
```

### Arguments

x	Matrix with sample in column and features in row
y	Matrix is compared to x.
dim	number of retained dimensions in the comparison. Defaults to all.

### Value

Ratio of projected variance to total variance

### Author(s)

Edgar Zanella Alvarenga

### References

Singhal, A. and Seborg, D. E. (2005), Clustering multivariate time-series data. *J. Chemometrics*, 19: 427-438. doi: 10.1002/cem.945

### Examples

```
c1 <- matrix(rnorm(16),nrow=4)
c2 <- matrix(rnorm(16),nrow=4)
pcasf(c1, c2)
```



---

plotcn	<i>plot PMD KEGG network for certain compounds and output network average distance and degree</i>
--------	---

---

**Description**

plot PMD KEGG network for certain compounds and output network average distance and degree

**Usage**

```
plotcn(formula, name, pmd)
```

**Arguments**

formula	Chemical formula
name	Compound name
pmd	specific paired mass distances

**Examples**

```
plotcn('C6H12O6', 'Glucose', c(2.016, 14.016, 15.995))
```

---

plotpaired	<i>Plot the mass pairs and high frequency mass distances</i>
------------	--

---

**Description**

Plot the mass pairs and high frequency mass distances

**Usage**

```
plotpaired(list, index = NULL, ...)
```

**Arguments**

list	a list from getpaired function
index	index for PMD value
...	other parameters for plot function

**See Also**

[getpaired](#), [globalstd](#)

**Examples**

```
data(spmein vivo)  
pmd <- getpaired(spmein vivo)  
plotpaired(pmd)
```

plotrtg *Plot the retention time group*

---

**Description**

Plot the retention time group

**Usage**

```
plotrtg(list, ...)
```

**Arguments**

`list` a list from `getpaired` function  
`...` other parameters for plot function

**See Also**

[getpaired](#), [globalstd](#)

**Examples**

```
data(spmein vivo)  
pmd <- getpaired(spmein vivo)  
plotrtg(pmd)
```

---

plotsda *Plot the specific structure directed analysis(SDA) groups*

---

**Description**

Plot the specific structure directed analysis(SDA) groups

**Usage**

```
plotsda(list, ...)
```

**Arguments**

`list` a list from `getpmd` function  
`...` other parameters for plot function

**See Also**

[getstd](#), [globalstd](#), [plotstd](#), [plotpaired](#), [plotstdrt](#)

**Examples**

```
data(spmein vivo)
re <- getpmd(spmein vivo, pmd=78.9)
plotsda(re)
```

---

plotstd	<i>Plot the std mass from GlobalStd algorithm</i>
---------	---

---

**Description**

Plot the std mass from GlobalStd algorithm

**Usage**

```
plotstd(list)
```

**Arguments**

list            a list from getstd function

**See Also**

[getstd](#), [globalstd](#)

**Examples**

```
data(spmein vivo)
pmd <- getpaired(spmein vivo)
std <- getstd(pmd)
plotstd(std)
```

---

plotstdrt	<i>Plot the std mass from GlobalStd algorithm in certain retention time groups</i>
-----------	--

---

**Description**

Plot the std mass from GlobalStd algorithm in certain retention time groups

**Usage**

```
plotstdrt(list, rtcluster, ...)
```

**Arguments**

`list` a list from `getstd` function  
`rtcluster` retention time group index  
`...` other parameters for plot function

**See Also**

[getstd](#), [globalstd](#), [plotstd](#), [plotpaired](#), [plotstdsda](#)

**Examples**

```
data(spmein vivo)
pmd <- getpaired(spmein vivo)
std <- getstd(pmd)
plotstdrt(std, rtcluster = 6)
```

---

<code>plotstdsda</code>	<i>Plot the std mass from GlobalStd algorithm in structure directed analysis(SDA) groups</i>
-------------------------	--

---

**Description**

Plot the std mass from GlobalStd algorithm in structure directed analysis(SDA) groups

**Usage**

```
plotstdsda(list, index = NULL, ...)
```

**Arguments**

`list` a list from `getsda` function  
`index` index for PMD value  
`...` other parameters for plot function

**See Also**

[getstd](#), [globalstd](#), [plotstd](#), [plotpaired](#), [plotstdrt](#)

**Examples**

```
data(spmein vivo)
re <- globalstd(spmein vivo, sda=TRUE)
plotstdsda(re)
```

---

pmdanno	<i>Perform MS/MS pmd annotation for mgf file</i>
---------	--

---

**Description**

Perform MS/MS pmd annotation for mgf file

**Usage**

```
pmdanno(file, db = NULL, ppm = 10, prems = 1.1, pmc = 0.6, scutoff = 0.1)
```

**Arguments**

file	mgf file generated from MS/MS data
db	database could be list object from 'getms2pmd'
ppm	mass accuracy, default 10
prems	precursor mass range, default 1.1 to include M+H or M-H
pmc	pmd length percentage cutoff for annotation. 0.6(default) means 60 percentage of the pmds in your sample could be found in certain compound pmd database
scutoff	relative intensity cutoff for input spectra for pmd analysis, default 0.1

**Value**

list with MSMS annotation results

---

runPMD	<i>Shiny application for PMD analysis</i>
--------	---

---

**Description**

Shiny application for PMD analysis

**Usage**

```
runPMD()
```

---

runPMDnet	<i>Shiny application for PMD network analysis</i>
-----------	---

---

**Description**

Shiny application for PMD network analysis

**Usage**

```
runPMDnet()
```

---

sda	<i>A dataset containing common Paired mass distances of substructure, ions replacements, and reaction</i>
-----	---

---

**Description**

A dataset containing common Paired mass distances of substructure, ions replacements, and reaction

**Usage**

```
data(sda)
```

**Format**

A data frame with 94 rows and 4 variables:

**PMD** Paired mass distances

**origin** potential sources

**Ref.** references

**mode** positive, negative or both mode to find corresponding PMDs

---

spmein vivo	<i>A peaks list dataset containing 9 samples from 3 fish with triplicates samples for each fish from LC-MS.</i>
-------------	---

---

**Description**

A peaks list dataset containing 9 samples from 3 fish with triplicates samples for each fish from LC-MS.

**Usage**

```
data(spmein vivo)
```

**Format**

A list with 4 variables from 1459 LC-MS peaks:

**mz** mass to charge ratios

**rt** retention time

**data** intensity matrix

**group** group information

# Index

## \* datasets

- hmdb, 14
- keggrall, 15
- omics, 15
- sda, 22
- spmein vivo, 22

sda, 22

spmein vivo, 22

- getcda, 2
- getchain, 3
- getcluster, 4
- getcorcluster, 5
- getms2pmd, 5
- getmspmd, 6
- getpaired, 4, 6, 8, 10–12, 14, 17, 18
- getpmd, 7, 10
- getposneg, 8
- getrda, 3, 8, 9, 10
- getreact, 10
- getsda, 3, 7–10, 11, 12, 14
- getstd, 4, 7, 8, 10, 11, 12, 14, 18–20
- gettarget, 13
- globalstd, 13, 17–20

hmdb, 14

keggrall, 15

omics, 15

- pcasf, 16
- plotcn, 17
- plotpaired, 7, 11, 17, 18, 20
- plotrtg, 18
- plotsda, 18
- plotstd, 4, 12, 14, 18, 19, 20
- plotstdrt, 14, 18, 19, 20
- plotstdsda, 14, 20, 20
- pmdanno, 21

runPMD, 21

runPMDnet, 21